

Screening of 33 Medicinal Plants for the Microelements Content

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Abstract

The microelements content of 33 medicinal plants was analyzed. The analysed microelements were: Iron, copper, zinc, manganese, cobalt, chromium, nickel, cadmium and lead. Mineral contents were determined by flame atomic absorption spectrometry (F-AAS) with high resolution continuum source ContrAA 300 spectrometer. The contents in microelements for analysed samples were in range: 18.1 ppm (*Symphytum officinale*) - 1.4 ppm (*Rhamnus frangula*), for Copper; 26,2 ppm (*Valeriana officinalis*) - 4,3 ppm (*Rhamnus frangula*), for Zinc; 214 ppm (*Violae tricoloris herba*) - 18 ppm (*Equisetum arvense*), for Manganese; 826 ppm (*Calendula officinalis*) - 23 ppm (*Rhamnus frangula*), for Iron. The microelements contents (Cu, Zn, Mn, Fe, Co, Cr, Ni, Cd and Pb) have grouped the analyzed medicinal plants in two main clusters. First main cluster was formed by other two groups.

Keywords: Cooper, iron, manganese, microelements, medicinal plants, Zinc

1. Introduction

Organic or inorganic salts are commonly present in foods as natural components, ingredients or additives. The latter are added to foods for a huge number of functional purposes. For instance, some of these act as flavouring, antimicrobial or pH control agents, and others as antioxidants and emulsifiers [1].

Mineral deficiency is usually caused by a low mineral content in the diet when rapid body growth is occurring and/or when there is poor absorption of minerals from the diet [2].

In this way, not only must the absolute amounts of minerals be increased in the edible portions of foods, but these minerals must also be in forms that are bioavailable for organism.

The metals contents of plants are variable, due to the factors like differences between the plants species, geographical area, conditions of drying process. Metals contents in soil are a great

importance for their effect on animals and humans, through the biologic chain: soil – plant – feed and food. The accessibility of metals for plants depends on soil reaction, mineral colloids, soil humidity, microbiological activity and organic matter content [3].

Zinc is a constituent of about 300 enzymes and proteins that participate in all major metabolic processes. As an essential trace element Zinc can impair vital function either by deficiency or excess. Dietary reference values for Zinc suggested by various sources (WHO-1996) range from 9.4 to 11.0 mg/day for male adults and from 6.5 to 8.0 mg/day for female adults.

Manganese is one of the essential microelements for plants, animals and humans. It is both a constituent and an activator of several enzymes and proteins in plant, animal and humans, and has around 20 identified functions [4].

Iron deficiency anaemia is a global nutritional problem, affecting nearly 1.78 billion people of which 358 million are from the developing world (WHO, 1998). Iron deficiency has a massive economic cost,

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adding to the burden on the health system, affecting cognitive performance of children and reducing adult productivity. The World Bank, WHO and Harvard University, list iron deficiency anemia as entailing a higher overall cost than any other disease except tuberculosis [5].

Also, Iron (Fe) is recognized as one of the most important trace elements for animal growth [6].

To prevent and treat iron deficiency, exogenous inorganic iron supplementation was popular in the feed industry. Iron sources from feed ingredients and supplements vary widely in their bioavailability to the animal [7].

Copper is one of several trace heavy metals that are essential to life, despite being as inherently toxic as nonessential trace heavy metals (Cadmium, Lead). NRC (1994) requirement of Cu for broiler chickens is only 8 mg/kg. When included at higher dietary levels, so-called pharmacological levels, this element has found to be effective for lipid metabolism [8].

Lead is not an essential element for life and it is very toxic for the nervous system and the kidneys. **Chromium** is involved in insulin function.

The goal of this paper was to analyse the microelements content for 33 medicinal plants (herbs). The analysed microelements were: zinc, manganese, iron, copper, cobalt, chromium, cadmium, lead and nickel.

2. Materials and methods

Medicinal plants or parts of them are used successfully in treatment of different kind of diseases for those therapeutically properties.

Medicinal plants samples preparation

Thirty three medicinal plants samples were collected of Romanian markets. The analysed medicinal plants are: *Achillea millefolium*, *Arnica montana*, *Artemisia absinthium*, *Calendula officinalis*, *Capsella bursa pastoris*, *Chelidonium majus*, *Crataegus monogyna*, *Cynara scolymus*, *Echinaceae herba*, *Epilobium montanum*, *Equisetum arvense*, *Galium mollugo*, *Hippophae rhamnoides*, *Hypericum perforatum*, *Malva silvestris*, *Matricaria chamomilla*, *Melissae Folium*, *Mentha piperita*, *Phoeniculus*, *Pinus*, *Plantago major*, *Poligonum aviculare*, *Rhamnus*

frangula, *Rosa canina*, *Salix babylonica*, *Salvia officinalis*, *Symphytum officinale*, *Taraxacum officinale*, *Thy mi herba*, *Tilia platyphyllos*, *Urtica dioica*, *Valeriana officinalis*, *Violae tricoloris herba*.

The metals content from medicinal plants samples were analysed after dry burning of 10g in the quartz capsules at 650°C for 4 hours. After complete burning a nitric acid 0,5 N solution was added up to 50 ml. The solution obtained were used for total metals contents determination by flame atomic absorption spectrometry (F-AAS) with high-resolution continuum source.

Reagents

The standard solutions (1000 mg / l) were analytical grade from Riedel de Haen (Germany). The nitric acid 65% solution used was of ultra pure grade (Merck, Germany). All solutions were prepared using deionised water.

Metals content determination

Analysis of metals content was made with ContrAA-300, Analytik-Jena device, by flame atomic absorption spectrometry (FAAS) in air/acetylene flame. The device working parameters (air, acetylene, optics and electronics) were adjusted for maximum absorption for each element. Acetylene was of 99.99% purity. Under the optimum established parameters, standard calibration curves for metals were constructed by plotting absorbency against concentration [9]. In a definite range for each metal a good linearity was observed. The correlation coefficient for the calibration curves (r^2) ranged between 0.9745-0.9891. All analyses were made in triplicate and mean values were reported. All the values obtained for metals contents in analysed samples were calculated in mg/kg dry matter (ppm).

3. Results and discussion

The *Cooper*, *Zinc*, *Manganese*, *Iron*, *Cobalt*, *Nickel*, *Cadmium*, *Chromium*, and *Lead* concentrations in analyzed samples are presented in table 1.

Table 1. The concentrations in Cooper, Zinc, Manganese, Iron, Cobalt, Chromium, Nickel, Cadmium and Lead in analyzed samples

Specification	Cu ppm	Zn ppm	Mn ppm	Fe ppm	Co ppm	Cr ppm	Ni ppm	Cd ppm	Pb ppm
Arnica montana	8	22.8	68	220	0.4	2.4	12.2	0	1.1
Cynara scolymus	6.7	16.8	87	705	0	5.3	13.9	0	0.6
Hippophae rhamnoides	4.5	8.1	122	87	0	0.4	4	0	0
Thy mi herba	6.4	14.9	78	283	0.3	1.3	6.4	0	0.4
Equisetum arvense	3.5	10.7	18	160	0	0.2	1.1	0	0
Achillea millefolium	5.9	11.4	43	152	0	0.3	5.7	0	0.5
Rhamnus frangula	1.4	4.3	29	23	0	0.1	0.5	0	0.6
Echinaceae herba	4.2	11.4	133	131	0.4	0.5	4.8	0	0
Phoeniculus	9.6	15.7	29	135	0	0.5	0.9	0	0
Calendula officinalis	10.5	14.8	59	826	1.1	5.7	3.6	0	0
Rosa canina	1.9	4.5	42	78	0	0	0.9	0	0
Mentha piperita	8	11.2	77	147	0	0.2	1.8	0	0
Matricaria chamomilla	7.7	17.2	65	244	0	1.2	3.9	0	0.6
Malva silvestris	4.4	14.9	31	310	0	0.8	3.1	0	0.6
Crataegus monogyna	7.6	16.3	49	255	0	0.2	0.7	0	0.5
Taraxacum officinale	5.5	13.5	33	245	0	0.8	1.1	0	0
Artemisia absinthium	5.3	12	59	629	0.3	2.9	5.4	0	1
Pinus	3.1	13.7	62	62	0	0.2	7	0	0.1
Plantago major	6.2	14	48	275	0	0.4	0.5	0	0
Epilobium montanum	4.9	12.9	60	84	0	0	2.1	0	0.2
Melissae Folium	5.6	11.2	50	185	0	0.4	4	0	0
Chelidonium majus	10	17.4	30	287	0	1.1	0.4	0	1.1
Salvia officinalis	4.9	12.6	52	450	0.1	2.2	5.6	0	0.5
Salix babylonica	2.3	23	25	32	0.1	0	3.2	0	0.3
Galium mollugo	3.8	13	43	75	0	0.2	0.6	0	0.3
Hypericum perforatum	8.6	16.9	75	155	0.1	0.7	2.1	0	0.1
Symphytum officinale	18.1	12.6	32	339	0.1	1.1	5.8	0	0.2
Tilia platyphyllos	8	13.5	73	113	0	0	0.8	0	0
Capsella bursa pastoris	4.3	11.4	29	212	0	1	3.1	0	0
Violae tricoloris herba	3.5	21.2	214	633	0	2.6	8	0	0.6
Poligonum aviculare	6.1	18.3	60	511	0.3	4.1	2.3	0	1
Valeriana officinalis	9.2	26.2	94	773	0.9	4.4	5.7	2.8	15.7
Urtica dioica	5.7	11.7	28	133	0	0.3	0.6	0	0.1

The cases - dendrogram obtained using Cu, Zn, Mn, Fe, Pb, Cr and Ni like variables is presented in Figure 1:

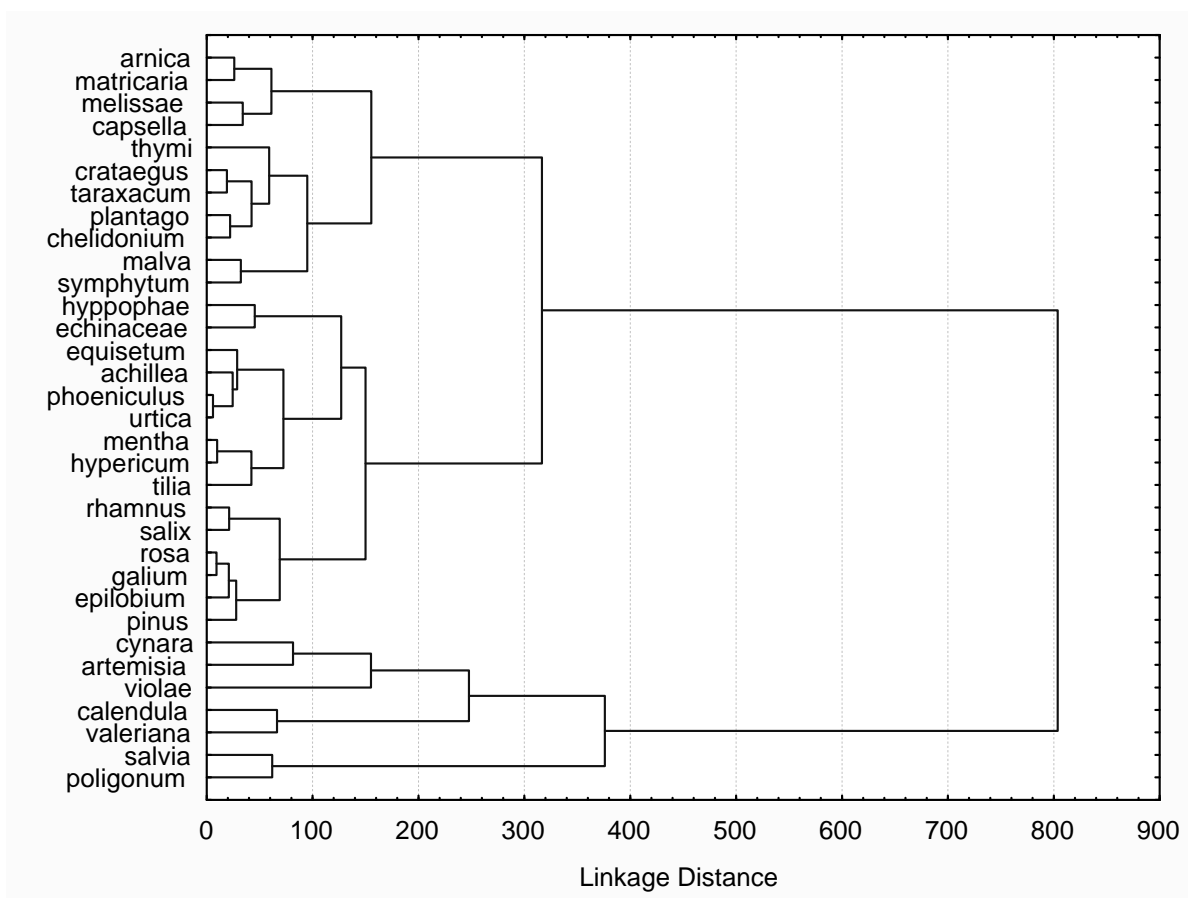


Figure 1 Clusters of medicinal plants using microelements

For cluster analysis it was used Statistica-6 software. The coefficients of matrix correlation of variables are presented in Table 2.

Table 2 The correlation coefficients of the microelements concentrations of medicinal plants

Microelements	Cu	Zn	Mn	Fe	Co	Cr	Ni	Cd	Pb
Cu	-								
Zn	0.30	-							
Mn	-0.08	0.26	-						
Fe	0.33	0.46	0.34	-					
Co	0.30	0.39	0.19	0.63	-				
Cr	0.29	0.47	0.29	0.93	0.70	-			
Ni	0.11	0.39	0.45	0.45	0.25	0.55	-		
Cd	0.17	0.46	0.16	0.41	0.54	0.36	0.11	-	
Pb	0.17	0.50	0.16	0.45	0.54	0.41	0.16	0.99	-

Marked correlations are significant at $p < 0.05$

The consistent correlation was found between Fe and Cr (0.93), Co and Cr (0.70) and Cd and Pb (0.99).

Copper deficiency in humans is a rare exception, and would not occur if Cu content were more than 2 mg in the daily diet. The national accepted limit for Cooper in tea is 50.0 mg/Kg (Ordinance 975/1998). For analysed samples Symphytum officinale has the highest content in Cooper (18.1 ppm), followed by Calendula officinalis (10.5

ppm), Chelidonium majus (10.0 ppm) and Phoeniculus (9.6 ppm). Rhamnus frangula has the smallest Cooper concentration (1.4 ppm).

The national accepted limit for **Zinc** in tea is 50.0 mg/kg (Ordinance 975/1998). The analysed samples contend Zinc in range 26.20 - 4.3 ppm. The smaller quantity was determinate for

Rhamnus frangula (4.3 ppm) and the higher for Valeriana officinalis (26.2 ppm).

Manganese In humans, the range between deficiency and toxicity of Mn is narrow. The recommended ESADDI values for adults range from 2 to 5 mg Mn/day [10]. The contents in Mn for analysed samples were in range 18.0 – 214.0 ppm. The low content was determined for Equisetum arvense (18.0 ppm) and the highest for Viola tricoloris herba (214.0 ppm).

Iron can not produce toxic effects in usual amounts. Nevertheless, the sum of published data suggests that Fe depletion beyond requirement may be hazardous [11,12].

Calendula officinalis has the greatest content in Iron (826.0 ppm), followed by Valeriana officinalis (773 ppm) and Cynara scolymus (705 ppm). Rhamnus frangula has the smallest content in Fe (23.0 ppm).

Nickel is an essential element for animal nutrition. Excessive soluble Ni compounds are hepatotoxic and nephrotoxic but as aerosols or dusts, insoluble Ni compounds or elemental Ni are very toxic (carcinogenic), justifying a lot of country imposed restricted limits, 0.05-1 mg/m³ [13].

Cynara scolymus has the great content in Nickel (13.9 ppm), followed by Arnica montana (12.2 ppm).

The higher quantity of **chromium** was determined for Calendula officinalis (5.7 ppm). For Tilia platyphyllos, Salix babylonica, Rosa canina and Epilobium montanum were not detected chromium content.

The effects of **Cadmium** are only partially understood. Cadmium interferes with the uptake, transport and use of different macro- and micronutrients, especially Iron and Zinc. Cadmium inhibit or activate a great enzymes number, like those rich in accessible sulfhydryl groups [14]. Also for a large number of medicinal herbs were not registered cobalt and cadmium contents.

Lead is not an essential element for life and it is very toxic for the nervous system and for the kidneys. The national accepted limit for Lead in tea is 5.0 mg/Kg (Ordinance 975/1998).

The microelements contents (Cu, Zn, Mn, Fe, Co, Cr, Ni, Cd and Pb) have grouped the analyzed medicinal plants in two main clusters. First main cluster was formed by other two groups. First group contained *Arnica montana*, *Matricaria chamomilla*, *Melissae Folium*, *Capsella bursa*

pastoris, *Thymi herba*, *Crataegus monogyna*, *Taraxacum officinale*, *Plantago major*, *Chelidonium majus*, *Malva silvestris*, and *Symphytum officinale*. All these medicinal plants had a medium content in Fe (185-339 ppm).

The second group of the first main cluster was formed by *Hippophae rhamnoides*, *Echinaceae herba*, *Equisetum arvense*, *Achillea millefolium*, *Phoeniculus*, *Urtica dioica*, *Mentha piperita*, *Hypericum perforatum*, *Tilia platyphyllos*, *Rhamnus frangula*, *Salix babylonica*, *Rosa canina*, *Galium mollugo*, *Epilobium montanum*, and *Pinus* characterized by small Fe concentrations, in range 87 -160 ppm. High Mn contents were determined for *Hippophae rhamnoides* (122 ppm) and *Echinaceae herba* (133 ppm). The smallest contents in Cu were obtained for *Rhamnus frangula*, *Salix babylonica*, *Rosa canina*, *Pinus*, *Epilobium montanum*, *Galium mollugo*, ranged 1.4 – 4.9 ppm. *Mentha piperita*, *Hypericum perforatum*, and *Tilia platyphyllos* had appropriate contents in Cu (8.0 – 8.6 ppm) and Mn (73 – 77 ppm).

The second main cluster grouped *Cynara scolymus*, *Artemisia absinthium*, *Viola tricoloris herba*, *Calendula officinalis*, *Valeriana officinalis*, *Salvia officinalis*, *Poligonum aviculare*. The highest content in Fe characterized this cluster (450-826 ppm).

4. Conclusions

The contents in microelements for analysed samples were in range: 18.1 ppm (Symphytum officinale) - 1.4 ppm (Rhamnus frangula), for Copper; 26,2 ppm (Valeriana officinalis) – 4,3 ppm (Rhamnus frangula), for Zinc; 214 ppm (Viola tricoloris herba) - 18 ppm (Equisetum arvense), for Manganese; 826 ppm (Calendula officinalis) - 23 ppm (Rhamnus frangula), for Iron. The microelements contents (Cu, Zn, Mn, Fe, Co, Cr, Ni, Cd and Pb) have grouped the analyzed medicinal plants in two main clusters. First main cluster was formed by other two groups.

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